Allosteric modulator identification from South African Natural Compounds Database (SANCDB) against SARS-CoV-2 M^{pro} protein in the presence of its evolutionary mutations

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South African Natural Compounds Database https://sancdb.rubi.ru.ac.za/ Hatherley et al. J Cheminform (2015) 7:29 ournal of **Chem**informatics DOI 10.1186/s13321-015-0080-8 DATABASE **Open Access** CrossMark SANCDB: a South African natural compound database Rowan Hatherley¹, David K Brown¹, Tho Diallo et al. J Cheminform (2021) 13:37 Kevin A Lobb^{1,2} and Özlem Tastan Bisho Journal of Cheminformatics https://doi.org/10.1186/s13321-021-00514-2 **Open Access RESEARCH ARTICLE** SANCDB: an update on South African natural compounds and their readily available analogs Bakary N'tji Diallo¹, Michael Glenister¹, Thommas M. Musyoka¹, Kevin Lobb^{1,2} and Özlem Tastan Bishop^{1*}



Identify allosteric inhibitors alternative to active site inhibitors.

How the behavior of allosteric inhibitors changes in the presence of the evolutionary mutations?

Allostery and mutations: Not commonly used biological phenomena in the early computational drug discovery stages.

Outline of study/talk

- **STEP 1:** Identification of M^{pro} mutations and their effects on the structure
- STEP 2: Identification of potential allosteric site(s) and effects of mutations on allosteric and active sites
- **STEP 3:** Identification of potential allosteric modulators (SANCDB) against reference structure
- STEP 4: Identification of functionally important residues and allosteric communication paths within protein-inhibitor complexes, and changes in the presence of mutations



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Article

Impact of Early Pandemic Stage Mutations on Molecular Dynamics of SARS-CoV-2 M^{pro}

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Novel dynamic residue network analysis approaches to study allosteric modulation: SARS-CoV-2 M^{pro} and its evolutionary mutations as a case study

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BASIC CONCEPTS

Two types of FDA approved drugs:

Orthosteric drugs Allosteric drugs

Definitions – Orthosteric site & drug

Orthosteric site: The functional site, e.g., active sites for enzymes, or protein-protein binding sites for receptors.

Orthosteric drugs bind to the functional site of a protein, compete with endogenous regulators and block the activity directly.

PROTEIN STRUCTURE

Scaffold to support and position active site

ACTIVE SITE

BINDING SITESCATALYTIC SITEBind and orient
substrate(s)Reduce chemical
activation energy



By Thomas Shafee - Own work, CC BY 4.0; https://commons.wikimedia.org/w/index.php?curid=45801894

Definitions – Allostery, allosteric site & drug

Two Greek words: "allos" - other/alternate & "stereo" - solid shapes.

Allosteric site: A site away from the orthosteric site but whose perturbation by binding an effector affects the conformation/function at the orthosteric site → allosteric communication.

Allosteric drugs modify the functional conformation and/or the active site of the target protein from a distance - allosteric site.



Perturbation is the key to allosteric behavior, and effectors that can cause perturbation are:



FDA approved drugs: Orthosteric versus allosteric

- The first discovery of allosteric systems > 60 years.
- The concept of using allosteric sites as drug targets is still not common.
- ~3700 FDA approved drugs.
- 19 of them allosteric drugs.
- 19 versus ~ 3681 drugs → hints at the difficulty of designing allosteric drugs.



Liu and Nussinov, 2016; https://doi.org/10.1371/journal.pcbi.1004966

Difficulty is due to:

- Identification of allosteric sites (shallow, cryptic pockets ...) serendipity
- Demonstration of the effects of allosteric modulators
 - inhibitor
 - activator

Allosteric drugs have benefits over orthosteric drugs:

 ✓ Allosteric sites are less conserved compared to active sites, therefore allosteric modulators are highly specific, hence may be less toxic to host.

Allosteric drugs versus orthosteric drugs: Specificity - toxicity



Nussinov and Tsai, Current Pharmaceutical Design, 2012

Allosteric drugs have benefits over orthosteric drugs:

✓ Unlike orthosteric drugs that compete with the substrate and cofactors, allosteric drugs can be active even in the presence of the native substrates, and thus reduce the chances of pathogen developing resistance by increasing the substrate concentrations and/or mutations.

Allosteric drugs versus orthosteric drugs: Resistance



Nussinov and Tsai, Current Pharmaceutical Design, 2012

Resistance \rightarrow brings us to other biological phenomenon:



NONSYNONYMOUS SUBSTITUTION a nucleotide mutation that alters the amino acid sequence of a protein

SYNONYMOUS SUBSTITUTIONS

a nucleotide mutation that does not alter amino acid sequences

MISSENSE MUTATIONS

mutations in a single nucleotide that result in the substitution of a different amino acid, resulting in a change to the protein encoded.

NONSENSE MUTATIONS

a mutation in the DNA sequence causes a protein to terminate prematurely by changing the original amino acid to a stop codon.

STEP 1: Identification of mutations and effects on the reference M^{pro} protein structure

Main Protease - M^{pro}

- 3C-like protease
- a conserved protein present in all members of the *Coronavirinae* subfamily
- does not have a human homolog good drug target
- functions as a homodimer
- Each monomer is call "protomer"
- SARS-CoV M^{pro} homolog
 - Homo-dimerization plays an important role in the catalytic activity of M^{pro}
 - Only one of the dimers was shown to be active at a time in SARS-CoV M^{pro}



Main Protease - M^{pro}

- comprises three domains (I-III)
- chymotrypsin-like domain I (residues 10-99)
- picornavirus 3C-protease like domain II (residues 100-182)
- domain III (helical domain) (residues 198-303)



form a hydrophobic substrate binding site, with catalytic residues HIS41 and CYS145

connected to domain II by a 15 residue linker loop, and was shown to regulate enzymatic activity in SARS-CoV.

Main Protease - M^{pro}

- The majority of the dimer contact interface is a result of interactions present between domain II (protomer 1) and the N-finger (protomer 2).
- In the same manner, the N-finger from protomer 1 contacts domain II from protomer 2.
- Dimer stability!



Reference sequence/structure: PDB ID: 5RFV 50 distinct mutant sequences were filtered: GISAID

GISAID ID Mutation **GISAID ID** Mutation **GISAID ID** Mutation A7V EPI_ISL_425284* T196M EPI_ISL_425319* A116V EPI_ISL_4244703 EPI_ISL_420422 G15D EPI_ISL_421005* P108S EPI_ISL_423642 T201A EPI_ISL_420181 G15S EPI_ISL_422860* EPI_ISL_419256* L220F A129V EPI_ISL_425242 G15S.D48E EPI_ISL_420579 P132L EPI_ISL_421506 L232F EPI_ISL_423772* M17I EPI_ISL_425655 T135I EPI_ISL_425235 A234V EPI_ISL_425342 V20L I136V K236R EPI_ISL_420182 EPI_ISL_426097* T45I N151D Y237H EPI_ISL_421312* EPI_ISL_420510* EPI_ISL_416720* EPI_ISL_425839* M49I EPI_ISL_415503 V157I EPI_ISL_425886* D248E V157L EPI_ISL_418269* R60C EPI_ISL_426028 EPI_ISL_418075 A255V EPI_ISL_420306 K61R EPI_ISL_417413 C160S EPI_ISL_422919 I259T EPI_ISL_421763* A70T EPI_ISL_418082 A173V EPI_ISL_423725 A260V G71S EPI_ISL_413021 EPI_ISL_423288 P184S EPI_ISL_425498* V261A EPI_ISL_415643 L89F EPI_ISL_420241* P184L EPI_ISL_421380* A266V EPI_ISL_420059 K90R EPI_ISL_419710* A191V,L220F EPI_ISL_420610* N274D EPI_ISL_419756 P99L EPI_ISL_415610* A193V EPI_ISL_425643 R279C EPI_ISL_425132 Y101C EPI_ISL_421515* EPI_ISL_422184* S301L T198I EPI_ISL_419984* R105H EPI_ISL_423007 T190I

Table 1: List of missense mutations in SARS-CoV-2 M^{pro}. Samples with noticeably large differences in C_{α} RMSD from the reference protease are marked with an asterisk.



D48E variant (from sample EPI ISL 425242) lead to a novel "TSEEMLN"" motif at the substrate binding flap, which may have an impact on substrate binding affinity or even specificity.

Figure 1: Mapping of the positions showing unique mutations from the reference M^{pro} sequence. For clarity, domains (I-III) are coloured (red, blue and orange respectively) only for one of the monomers, while the other is represented as a grey surface. The domain linker region is in green and the N-finger is in cyan. The size of the labels denotes the number of unique mutations recorded at that position.

Estimation of the protein backbone flexibility from MD using C_{alpha} RMSD



Figure 3: (A) Violin plots of C_{α} RMSD values for the reference (in grey) and the mutant (coloured in blue) M^{pro}, showing the 25th, 50th and 75th percentiles in dotted lines inside the kernel density plots. Distributions are scaled by area, and have been sorted by the KDE D_J distance (shown above each distribution) computed between the each sample and the reference protease.

- A7V occurs on the N-finger, which is a critical region for M^{pro} dimer stability.
- M17I occurs on an internal loop that connects a strand to a helix in domain I.
- A70T occurs on solvent-exposed loop in the same domain.
- A116V occurs in a buried strand within domain II.

N-finger can adopt a range of equilibrium conformations, with some showing protomer symmetries and others not



Figure 5: Kernel density distributions of RMSD values for the N-finger region across the mutant and reference protease complexes. The violin plots are split in two for each protein sample, showing the RMSD values for chains A (in blue) and B (in red). The tips of the distributions mark the minimum and maximum values for both chains combined in each complex. Samples have been sorted by increasing median distance between the chains, also shown (in Å) at the top of each sample distribution.

Interprotomer distances are estimated using COM distance



Figure 8: Distributions of interprotomer COM distances across samples, arranged in ascending order of the KDE d_J . The reference protease is in grey while the mutants are coloured from yellow to red, in increasing order of distance from the reference KDE.

Inter-domain angles in each protomer of Mpro are calculated



05 9 6 0 N N M 0 4 8 0 50 6 0110 0100100100000 $\infty \infty$ 0 L-MMUOOH000MM 040 MUJJOUOJONH 0 NO 2 S 0 0 0014000404000000000000000040000 ote 20.20 22 21 0000000 MM000HN00 0 10 8 0 LO 0 m 4 0 LO NMAOM® 0 10 OH **NH** NN SSS Р 0 0 Ч Ч Р 0 Samples

Figure 10: Kernel density distributions of inter-domain angles (domains I-II-III) across the mutant and reference protease complexes. The violin plots are split in two for each protein, showing the inter-domain angles for chains A (in blue) and B (in red). The tips of the distributions mark the minimum and maximum values for both chains combined in each protein complex.

STEP 2: Identification of potential allosteric site and effects of mutations on allosteric and active site

Dual allosteric pockets coinciding with various stabilizing and functional components of the substrate binding pocket is identified



Figure 12: Pocket detection using combined predictions from FTMap and PyVOL. FTMap probes are shown as stick figure representations, while those from PyVOL are shown as surfaces. The protomers are depicted as cartoon representations, in grey and light orange.

An asymmetry in compaction between substrate binding pockets coming from each protomer in each sample is observed



Figure 13: Kernel density distributions of R_g values for the substrate binding site from each protomer of M^{pro} , arranged in ascending order of difference in median from each chain. The differences, shown at the top are in nanometres. Chain A values are in red while chain B values are in blue. The maxima and minima are across protomers. Quartiles for each binding site are shown as dotted lines.

The interprotomer (allosteric) pockets may play an important role in affecting the degree of compaction of the binding cavity and vice-versa



Figure 15: Kernel density distributions of R_g values across samples for the mirrored interfacial (and potentially allosteric) pockets. The differences, shown at the top are in nanometres.

STEP 3: Identification of potential allosteric modulators

SANCDB compounds have relatively different behavior in the M^{pro} protein of SARS-CoV-2 and of HCoV-OC43



Potential allosteric modulators in the presence of mutations



Six allosteric modulators were stable only on the 3 mutant systems

Isolate	Position	302	303	467	468	469	630	Consensus	Isolate	Position	302	303	467	468	469	630	Consensus
EPI_ISL_413021	G71S	X		X	X	X		4	EPI_ISL_421506	L232F	х	X		X	X		4
EPI_ISL_415503	V157I		X	X				2	EPI_ISL_421515	T198I			X	x	X		3
EPI_ISL_415610	A193V	X	X	X	X	x		5	EPI_ISL_421763	А70Т			X	x	X	X	4
EPI_ISL_415643	L89F		X	X	X			3	EPI_ISL_422184	S301L				x	X	X	3
EPI_ISL_416720	Y237H			X	X	X		3	EPI_ISL_422860	A129V		X			X	X	3
EPI_ISL_417413	C160S				X	x		2	EPI_ISL_422919	I259T			X	x		X	3
EPI_ISL_418075	A255V				X		X	2	EPI_ISL_423007	T190I	х			x	X	X	4
EPI_ISL_418082	A173V	X	X	X	X	X	X	6	EPI_ISL_423288	P184S	x			x	X	X	4
EPI_ISL_418269	R60C	X			X	X		2	EPI_ISL_423642	T201A			X	X	X	X	4
EPI_ISL_419256	L220F				X	X	X	3	EPI_ISL_423725	A260V		X	X	X	X	X	5
EPI_ISL_419710	A191V, L220F			X	X	x	X	4	EPI_ISL_423772	M17I			X	x	X	X	4
EPI_ISL_419756	P99L				X	x	X	3	EPI_ISL_424470	T196M	х			x	X	X	4
EPI_ISL_419984	R105H	X			X	X	X	4	EPI_ISL_425132	Y101C	х		X	X	X		4
EPI_ISL_420059	K90R	X	X			x		3	EPI_ISL_425235	A234V	х				X	X	3
EPI_ISL_420181	G15S	X	X	X		x		4	EPI_ISL_425242	G15S, D48E		X	X	x	X	X	5
EPI_ISL_420182	I136V			X	X	x		3	EPI_ISL_425284	A116V		X	X	x		X	4
EPI_ISL_420241	P184L				X		X	2	EPI_ISL_425319	A7V	х	X			X		3
EPI_ISL_420306	K61R		X		X	X		3	EPI_ISL_425342	V20L		X	X	X	X		4
EPI_ISL_420422	G15D	X		х	х	х		4	EPI_ISL_425498	V261A			х	x	X		3
EPI_ISL_420510	N151D				X	X		2	EPI_ISL_425643	R279C	х	X	X	X	X	X	6
EPI_ISL_420579	P132L	X		Х			X	3	EPI_ISL_425655	T135I			X	x	X	X	4
EPI_ISL_420610	N274D	X	X	X	X	X	X	6	EPI_ISL_425839	M49I		X	X	x	X	X	5
EPI_ISL_421005	P108S		X		X		X	3	EPI_ISL_425886	D248E			X	X	X	X	4
EPI_ISL_421312	T45I	X	X		X		X	4	EPI_ISL_426028	V157L		X		X	X	X	4
EPI_ISL_421380	A266V		X	X	X	X	X	5	EPI_ISL_426097	K236R	X	X	X	X	X		5
											20) 22	29	43	3 41	30	

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SANCDB	Search Tools Documentation Cite Us	About Us - Download Database		
Search Q SEARCH OPTIONS	SANC00468			
© SMILES ∄ Structure	Entry name:	4-Hydroxy-3-methoxybenzoic acid		
PropertiesSource Organism	Formula:	C ₈ H ₈ O ₄		SMILES:
 Classification Use 	Molecular mass:	168.15	HO	Search PubChem SMILES
▲ Author	ChEMBL ID:	CHEMBL120568	~ ОН	Scaffold SMILES:
c Advanced	ChemSpider 1D:	8155	2D Image 3D View	
	PubChem ID:	CID: 8468	View analogs	Peferences
	CAS No.	121-34-6		Koorbanally et al. (2004) Bufadienolides t Koorbanally et al. (2005) A novel homoise

- lides from Drimia robusta and Urginea epigea (Hyacinthaceae)
- omoisoflavonoid from Drimia delagoensis (Urgineoideae: Hyacinthaceae)

Classifications

- Aromatic acid
- Benzene and substituted derivatives (Classyfire)

Other Names

- 4-Hydroxy-3-Methoxybenzoic Acid
- Benzoic Acid, 4-Hydroxy-3-Methoxy-
- Vanillic Acid
- 2-Methoxy-4-Carboxyphenol
- 3-Methoxy-4-Hydroxybenzoic Acid Nsc 3987
- Nsc 674322
- Va
- M-Methoxy-P-Hydroxy-Benzoic Acid

Source Organisms

- Drimia robusta
- Drimia delagoensis

Compound Uses None Recorded

STEP 4: Hub (centrality) residues and allosteric communication paths (if any) and changes in the presence of **mutations**



Analysis of missense mutations: Proposed protocol











FIGURE 3.3

The node at the center of the cluster in the upper right would have a high degree centrality, even though it is far from the dense center of the network.

- betweenness centrality (BC),
- closeness centrality (CC),
- degree centrality (DC),
- eigencentrality (EC),
- katz centrality (KC)

The averaged *BC* metric is defined as how often a residue is traversed along the shortest paths connecting every other residue pairs.

Averaged *CC* of a residue is calculated as the reciprocal of the average number of the shortest paths linking a residue n and all other residues in the network

DC defines the number of neighboring nodes (the local connectivity) around a given node.

EC measures the high centrality given to high degree residue, or to a residue that is connected to other high degree residues.

KC measures the relative degree of influence of a residue i within connected residues in a network.

The term "centrality" is used as a measure of how central a residue is in the protein network, and several centrality metrics derived from the social sciences Bioinformatics, 33(17), 2017, 2768–2771 doi: 10.1093/bioinformatics/btx349 Advance Access Publication Date: 31 May 2017 Applications Note

OXFORD

Structural bioinformatics

MD-TASK: a software suite for analyzing molecular dynamics trajectories

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MDM-TASK-web: MD-TASK and MODE-TASK web server for analyzing protein dynamics



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A R T I C L E I N F O

ABSTRACT

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The web server, MDM-TASK-web, combines the MD-TASK and MODE-TASK software suites, which are aimed at the coarse-grained analysis of static and all-atom MD-simulated proteins, using a variety of non-conventional approaches, such as dynamic residue network analysis, perturbation-response scanning, dynamic cross-correlation, essential dynamics and normal mode analysis. Altogether, these tools allow for the exploration of protein dynamics at various levels of detail, spanning single residue pertur-



METHODOLOGY







Heat map for the potential hubs according to averaged *EC* metric for the reference and 50 mutant proteins in allosterically bound state to SANC00302 and SANC00468.



The communication path traced by averaged *EC* hubs, starting from the allosteric ligand towards the catalytic residue



- (A) M^{pro}-SANC00302 reference protein ligand complex. Allosteric modulator is in green.
- (B) (B) Mutant M^{pro} (G71S)-SANC00302 complex. Mutant (in purple) indicated by arrow.
- (C) C) M^{pro}- SANC00468 reference protein ligand complex. Allosteric modulator is in purple.
- (D) (D) Mutant M^{pro} (A173V)-SANC00468 complex.

Conclusion

Collectively, our approaches offer routes for novel rational drug discovery methods and provide computationally feasible platforms

Acknowledgement



https://covidrug-africaconsortium.rubi.ru.ac.za/

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